

ORIGINAL ARTICLE

PEER REVIEWED | OPEN ACCESS

Morphological and morphometric study on the early developmental stages of the Egyptian toad *Bufo regularis* Reuss

Alaa El-Din H. Sayed, Ekbal T. Wassif, Afaf I. Elballouz

ABSTRACT

Aims: This study aimed to examine the developmental changes in external morphology characters for some embryonic larval period of *Bufo regularis* from River Nile, Assuit, Egypt. **Methods:** Our experiment started when tadpoles began to feed. The adapted embryos were divided into three large tanks, each with 200 embryos. Collecting of samples started from feeding age every three days until end of metamorphosis (the absorption of tail). External measurements of the early developmental stages including body height (BH), body width (BW), eye diameter (ED), forelimb length (FLL), head-body length (H-BL), hind limb length (HLL), internarial distance (IND), interorbital distance (IOD), nostril diameter (ND), distance between narial and orbital (NOD), tail length (TAL), tail height (TH), total length (TL), tail muscle height (TMH) and body weight (BWE) were recorded. **Results:** The morphological description reported for each stage indicated many differences between the different developmental stages. **Conclusion:** This study may be considered as morphological key for the different developmental stages of the Egyptian toad tadpoles (*Bufo regularis*).

Keywords: *Bufo regularis*, Egyptian toad, Morphology, Tadpoles

How to cite this article

Sayed AH, Wassif ET, Elballouz AI. Morphological and morphometric study on the early developmental stages of the Egyptian toad *Bufo regularis* Reuss. Edorium J Anat Embryo 2016;3:1–14.

Article ID: 100009A04AS2016

doi:10.5348/A04-2016-9-OA-1

INTRODUCTION

Bufonidae represent one of the common anuran families distributed in all parts of the world except for Antarctica. About 300 species are included in the *Bufo* genus. The common species in Egypt are *Bufo regularis*, *Bufo viridis*, *Rana bedriagae* and *Ptychadena mascareniensis* [1]. The knowledge of tadpole morphology is a valuable tool for different biological studies, i.e. taxonomic, ecological, bio-geographic and faunistic registers. In addition, it had been reported that the knowledge of tadpole's morphological aspects can serve as a support to understand species preservation, and information that helps understand basic and derivative aspects of these species [2].

Anuran metamorphosis consists of an early period of larval growth (premetamorphosis), a subsequent interval of continued growth and development of some adult organs (prometamorphosis) and a final period when animals cease feeding, larval organs are resorbed, and final transformation to the juvenile frog occurs (climax) [3, 4].

Alaa El-Din H. Sayed¹, Ekbal T. Wassif¹, Afaf I. Elballouz^{1,2}
Affiliations: ¹Zoology department, Faculty of Science, Assiut University, 71516 Assiut, Egypt; ²Zoology department, Faculty of Science, Misurata University, Misurata, Libya.
Corresponding Author: Alaa El-Din Hamid Sayed, Zoology department, Faculty of Science, Assiut University, 71516 Assiut, Egypt; Email: alaa_h254@yahoo.com (A. H. Sayed)

Received: 21 October 2015
Accepted: 17 December 2015
Published: 29 February 2016

Two approaches have described the anurans ontogeny, the first one has named life cycle events as stages; embryonic, larval, metamorphic climax and juvenile/adult [5, 6]. The second approach has addressed ontogeny as metamorphosis that can be subdivided into three periods; premetamorphosis, prometamorphosis, and metamorphic climax [7–10]. Some studies investigated that the beginning of the larval period as when feeding begins [10] while others stated that the completion of hatching is the beginning [5, 6]. When comparing species, the morphological development at hatching differs between species [7, 8, 10] observed that the emergence of forelimb can be used as a mark of the beginning of metamorphic climax, where in some specialized anuran larvae, the initiation of mouthpart regression and forelimb emergence may be decoupled [6, 11].

It has been reported that the anuran metamorphosis is controlled by the hypothalamo-hypophyseal-thyroid axis involving actions of several hormones [12–14] such as thyrotrophic releasing hormone (TRH), thyroid hormones (TH), and prolactin [4, 15, 16]. Considerable flexibility has been observed in the rates of growth and development by several authors [17, 18].

Morphological characteristics are important marks to distinguishing between tadpoles of different species [19]. The external larval stages can also be identified on the basis of age, size and external morphological characters [4].

Only two works have been published on the external morphology of the early development of the tadpole of *Bufo regularis* by several authors [20, 21] who had suggested a generalized table of 66 stages covering the period from fertilized eggs to the completion of metamorphosis. In the present works the tadpoles of the Egyptian toad *Bufo regularis* will be re-described, The normal table that suggested by Sedra and Michael [20] are still the source of information about these species. The aim of the present study is to identify the different developmental stages of the Egyptian toad *Bufo regularis*; a step toward building a morphological and morphometric key for larval stages of the Egyptian toad (*Bufo regularis*). This search represented the first one of our series future papers about the development of *Bufo regularis* tadpoles.

MATERIALS AND METHODS

Specimen collection

Fertilization process and tadpoles care was done according to Sedra and Micheal [20]. Specimen collection was according to Sayed et al. [22].

Experimental Setup

Experimental setup was described in our previous publications [22, 23]. All water was changed daily.

Fifteen samples were taken and fixed in Bouin's solution for morphological preparations and morphometric measurements.

Morphology and Morphometric measurements

The staged tadpoles which fixed in Bouin's fluid were used for the morphological studies. The external morphological characters were recorded by using a binocular dissecting microscope fitted with ocular micrometer (NOVEL EDICALCO., LTD. XSZ-109 B) and photographed by the assist of a digital colored video camera (Sony, AVT Horn). All measurements were taken by the help of the ocular micrometer at known magnifications and Vernier calipers were used for the largest specimens (measured to the nearest 0.05 mm). The weight was measured using digital precision scales of one gram.

The external morphology of the different developmental stages were recorded including: Body Height (BH), Body Width (BW), Eye diameter (ED), Forelimb Length (FLL), Head- Body Length (H-BL), Hind Limb Length (HLL), Internarial Distance (IND), Interorbital Distance (IOD), Nostril Diameter (ND), Distance between Narial and orbital (NOD), Tail Length (TAL), Tail Height (TH), Total length (TL), Tail Muscle Height (TMH) and Body Weight (BWE). The body parts were based on the proposals of [24]. The external morphometric measurements were described in (Figure 1a–b).

Statistical analysis

The basic statistics, means, standard divisions and ranges were calculated. The pattern of variation was analyzed using one-way analysis of variance, using SPSS package [25] at the 0.05 significance level, a person's correlation was used.

Ethical statement

All experiments were carried out in accordance with the Egyptian laws and University guidelines for the care of experimental animals. Procedures of the current experiment have been approved by the Committee of the Faculty of Science of Assiut University, Egypt.

RESULTS

The important larval stages were differentiated on the basis of age, size and external morphological characters. Various stages of development and metamorphosis of *Bufo regularis* were divided into 13 stages. These were collected every three days from beginning of feeding to the end of the metamorphosis. This description was based on the normal table of [20]. The investigators described only the new structures appeared in each stage.

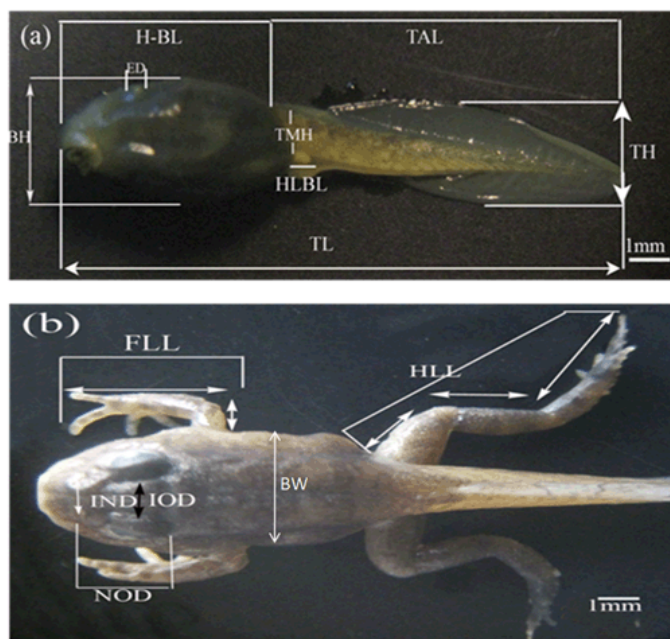


Figure 1: Morphometric measurements of *Bufo regularis* tadpoles.

Lateral view (stage 48); (b) Dorsal view (stage 64) Body Height (BH), Body Width (BW), Eye Diameter (ED), Forelimb Length (FLL), Head- Body Length (H-BL), Hind Limb Length (HLL), Internarial Distance (IND), Interorbital Distance (IOD), Distance between Narial and Orbital (NOD), Tail Length (TAL), Tail Height (TH), Total Length (TL), Tail Muscle Height (TMH). Scale bar = 1 mm.

Morphology

The important larval stages were differentiated on the basis of age, size and external morphological characters. Various stages of development and metamorphosis of *Bufo regularis* were divided into 13 stages. These were collected every three days from beginning of feeding to the end of the metamorphosis. This description was based on the normal table of [20]. The investigators described only the new structures appeared in each stage.

Stage 44: Start feeding stage (TL; 7.5–7.5 mm, BWE; 0.003–0.005 g).

At the beginning of feeding the body (head and abdomen) is oval in shape and pigmented dorsally, but ventrally there is no pigments. The dorsally located eyes are moderately large in size. The nares are oval in shape and positioned between the eyes and the oral disc. The protocercal tail is tapering and laterally compressed (Figure 2a–b). The pigmented tail musculature is poorly developed and gradually tapering from base to the sharp end. Unpigmented tail fin consisted of dorsal and ventral ones. They are slightly equal in size. The dorsal tail fin slightly smaller than the ventral one. Dorsal fin originates at body – tail junction and expanded slightly above the body. The lateral tubular vent is moderately large and directed posteriorly, entirely attached to ventral fin (Figure 2b).

Coiling intestine appeared as triple U-shape ventrally. The mouth is located on the anteroventral side of the head. The small oral disc is transversally elliptical and consists of upper and lower lip. These lips carry teeth (Figure 2a&b).

Stage 46: Appearance of the hind limb buds (TL; 8–10 mm, BWE; 0.0043–0.0068 g)

In this stage, hind limb bud was visible for the first time as a faintly circumscribed projection in the groove between the base of the tail and body wall, (Figures 2c and 6a). Horny labial tooth rows of upper and lower oral disc were well developed. The first indication of two narrow longitudinal strands of muscle fibers of the *M. rectus abdominis* were observed through the transparent skin of the abdomen. Intestine is more coiling. Melanophores began progressively spread ventrally. Segmentally arranged myotomes are noticed for the tail muscles (Figure 2c–d).

Stage 48: Length of hind limb bud was equal to one and half times of its diameter (TL 12.5–13 mm; BWE; 0.024–0.036 g)

Length of hind limb bud was equal to one and half times of its diameter (Figures 2e and 6 c). Horny teeth and sub marginal papillae very highly developed. Through the transparent skin the elongation of the two narrow longitudinal strands of muscle fibers of the *M. rectus abdominis* was observed (Figure 2f). The Spiral coiling of the intestine increased, central coil of intestine still lies on the same level as that of the surrounds. Eye lens was very distinct. Spreading of blood capillaries in the dorsal fin was seen (Figure 2e–f).

Ventral view of the tadpole showing a typical oral disk apparatus at this stage which consists of upper and lower labia, enclosing upper and lower serrated jaw sheath (horny beak). The lower serrated jaw sheath of the horny beak appeared as V shape and the upper one was crescent in shape. The upper and lower labial ridges had horny teeth. There are two rows of horny teeth in the upper lip; the anterior one is complete, while the posterior one is discontinuous medially. The lower lip has three continuous rows of horny teeth. The number and configuration of tooth rows was indicated by a labial tooth row formula (LTRF) that is 1: (1+1)/3 according to Sedra and Michael (1961). The labia were framed by lateral rows of marginal and sub marginal papillae (Figure 3).

Stage 52: Foot paddle stage (TL; 15.3–17.4 mm; BWE; 0.041–0.046 g)

Landmarks between parts of the hind limb were slightly distinguished. Distal end of hind limb buds were moderately flattened to form the foot paddle and slightly bent ventrally (Figures 2g and 6e). No interdigital indentation was noticed at the paddle margin. At the end of this stage first interdigital indentation were noticed. The fifth and fourth toe prominences were separated by a slight indentation at the margin of the foot paddle. Through the thin transparent skin well developed *M. rectus abdominis* and tail muscles were seen. Extremely

distinctive segmentally arranged myotomes were noticed of the tail muscle. Each myotome is completely enclosed in a sheet of fibrous connective tissue (myocomma). The myotomes are >-shaped, each having an upper and lower limbs; their enclosing myocomma was also of the same shape and well visible externally (Figure 2g–h).

Stage 54: Second indentation (TL; 17.9–19.2 mm, BWE; 0.054–0.05 g)

The hind limbs were well-differentiated into thigh, shaft and foot. Second indentation occurred on the ventral side behind the prominence of 4th toe, which marks the 3rd toe (Figure 2i and 6g). Muscle fibers of *M. rectus abdominis* were more distinct forming an elongated sheet on either side of the middle line; this muscle is broad posteriorly and thinner anteriorly. Tendinous intersections were clearly observed (Figure 2j). Highly developed sub marginal papillae of the oral disc were noticed. Central coil of intestine pushed its way inward, (Figure 2i–j).

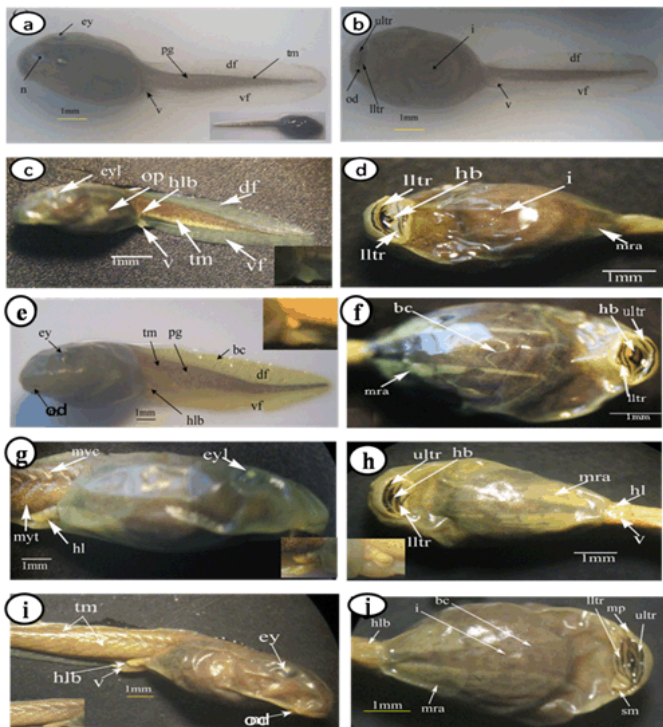


Figure 2: (a & b) Developmental embryonic stages of *Bufo regularis*, (c & d) Lateral and ventral view of stage 44 in situ showing the dorsal view. Lateral and ventral view of stage 46, in situ showing vent in (c) and oral disc and *M. rectus abdominis* in (d). (e & f) Lateral and ventral view of stage 48, in situ showing hind limb bud (e) (g & h) Lateral and ventral view of stage 52 in situ showing the foot paddle (g) and first indentation in (h) at the end of this stage. (i & j) Lateral and ventral view of stage 54, in situ showing the second indentation), bc = blood capillaries, df = dorsal fin, ey = eye, eyl = eye lens, hb = horny beak, hl = hind limb, hlb = hind limb bud, i = intestine, lltr = lower lip of teeth rows, mra = muscle rectus abdominis, mp = marginal papillae, myc = myocomma, myt = myotomes, n = nose, od = oral disc, op = operculum, pg = pigments, sm = sub marginal papillae, tm = tail muscle, ultr = upper lip of teeth rows, v = vent, vf = ventral fin and scale bar = 1 mm.

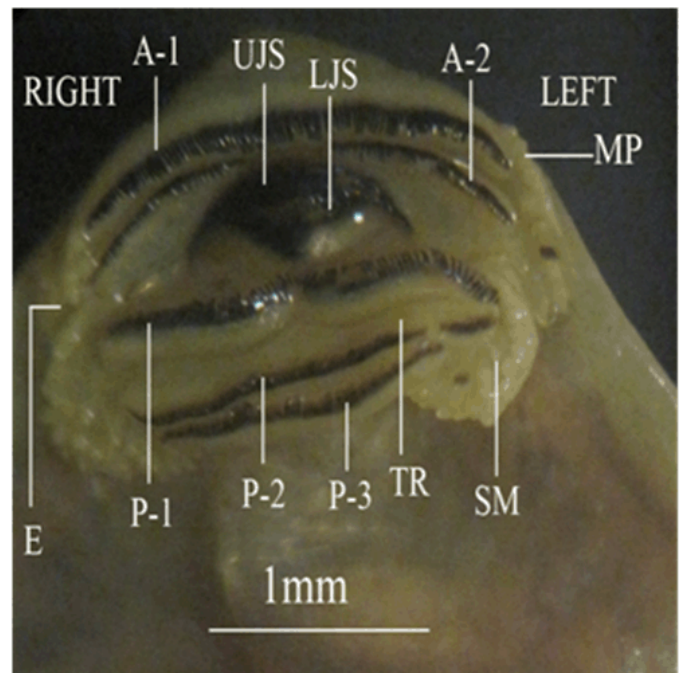


Figure 3: Ventral view of well developed oral apparatus of stage 48 in *B. regularis* with 1: (1+1)/3 tooth rows formula showing, A-1 and A-2 first and second anterior tooth rows notice the gap in the second row of the dorsal lip, lateral emargination of oral disc (E), lower jaw sheaths (LJS), marginal papillae (mp), P-1-3- first through third posterior tooth rows, Sub Marginal papillae (SM), tooth ridges for tooth row P-2 (TR), upper jaw sheath (UJS) and scale bar = 1 mm.

Stage 55: Third indentation (TL 17–18.3 mm, BWE; 0.081–0.087 g)

The margin of foot paddle became indented on ventral side, behind the prominence of 3rd toe which marks the prominence of 2nd toe. Pigments of melanophores were noticed on the thigh and shank of the hind limbs (Figures 4a and 6h). Tail musculature was strong, gradually tapering, almost reaching tail tip. Tail fins were moderately high, not extending above the body (Figure 4a–b).

Stage 56: Fourth indentation (TL; 21.5–22 mm, BWE; 0.094–0.097 g) Hind limbs were highly differentiated into thigh, shank and foot, pigmentation is spreading on the upper surface of the shank as well as on the toe prominences (Figure 4 c and 6i). The margin of foot paddle was indented behind the second toe demarcating the prominence of the first toe. All five toes were visible and separated from each other (Figure 4c). Through the transparent skin well developed forelimbs could be seen and were still concealed by the skin of the branchial chambers (Figure 4c & d).

Stage 57: Toe web ws more distinct (TL; 22.5–23, BWE; 0.133–0.138 g)

Landmarks between thigh, shank and foot were more differentiated and well distinct (Figure 4e); the margin of 5th toe web was directed toward the tip of 1st toe.

Pigmentation appeared in the 4th and 5th toes along the foot. Toes were longer and all toes were separated from each others. Prehallux appeared as a small protuberance close to the base of the first toe. The fourth toe was the longest. Web clearly developed between 2–3 and 3–4 toes. Instep carrying little tubercles some of which are enlarged at bases of the second, third, and fourth toes (Figure 7).

Muscle of *M. rectus abdominis* are divided into five pairs of transverse compartments by four pairs of tendinous intersections, the posterior three pairs of these compartments are highly developed as compared with the anterior ones, vent tube are highly distinct (Figure 4e–f).

Stage 58a: Appearance of toe pads (TL; 25–25.3 mm, BWE; 0.154–0.158 g)

The fourth toe was the largest and twice as long as the third toe. All toes were better differentiated and toe webs more distinct, the third and fifth finger were more or less equal to each other, fingers were stretched out in forelimb atrium which was still concealed by the skin of the operculum, horny teeth and jaws are still present. Muscle of *M. rectus abdominis* was still clearly indicated. Tendinous intersections were still obvious through skin of abdomen and replaced with muscle mass (Figure 4g–h and Figure 6j).

Stage 58b: Cloacal tail piece reduced (TL; 24 - 24.75 mm, BWE; 0.163–0.168 g)

Forelimbs were in process of eruption and visible through the skin (Figure 4i), elbows were pressing the overlying skin, palm well-built and is provided with a number of tubercles, vent tube disappear completely, cloacal tail piece reduced and tail darkening starts. The disappearance of teeth started (Figure 4i & j and 6k).

Stage 59: Both forelimbs emerge (TL; 27–27.3 mm, BWE; 0.15–0.16 g)

The left forelimb had passed out throughout the wide, spout-like opening of branchial chamber (Figure 5a). About 4–5 tubercles on the ventral surface of each finger are present, the fourth toe is the longest. One of the tubercles on palm is more prominent. Instep carrying little tubercles some of which are enlarged at bases of the second, third and fourth toes (Figure 5b), Right forelimb has passed out, thus no continuity between skin of body and limbs. Head is semicircular in outline and clearly broader than trunk, all horny teeth and lower horny jaw have fallen down. External nares were nearer to the tip of the snout than eye. External nares were opened at this stage. Warts were appeared (Figure 5a). Muscle of *M. rectus abdominis* were faintly indicated through the relatively thick and opaque skin of abdomen (Figure 5a&b and 6i).

Stage 64: Angle of mouth reached posterior border of the eye (TL; 20–23 mm, BWE; 0.153–0.156 g)

Tail decreased in size, dorsal and ventral lobes of caudal fin were highly reduced and represented by shallow apices (Figure 5c). Mouth is wider and its corners lie at the level of the posterior border of eye lens, the angle of

the mouth has reached posterior of the eye (Figure 8d). Rapid changes in the mouth and tail had been occurred. Linea Alba in the ventral side of the tadpole was noticed (Figure 5c&d and 6n).

Stage 66: Metamorphosed into toadlet (TL; 15.8–16 mm, BWE; 0.093–0.096 g)

Newly metamorphosed toadlet: Tail has disappeared and is only represented by a minute knob which is no longer visible in ventral aspect (Figure 5e). The corner of the wide mouth lies slightly behind the posterior border of the eye. Parotoid glands lie nearly behind the eyes. The animal had small size. Horny jaws were fallen. Linea alba

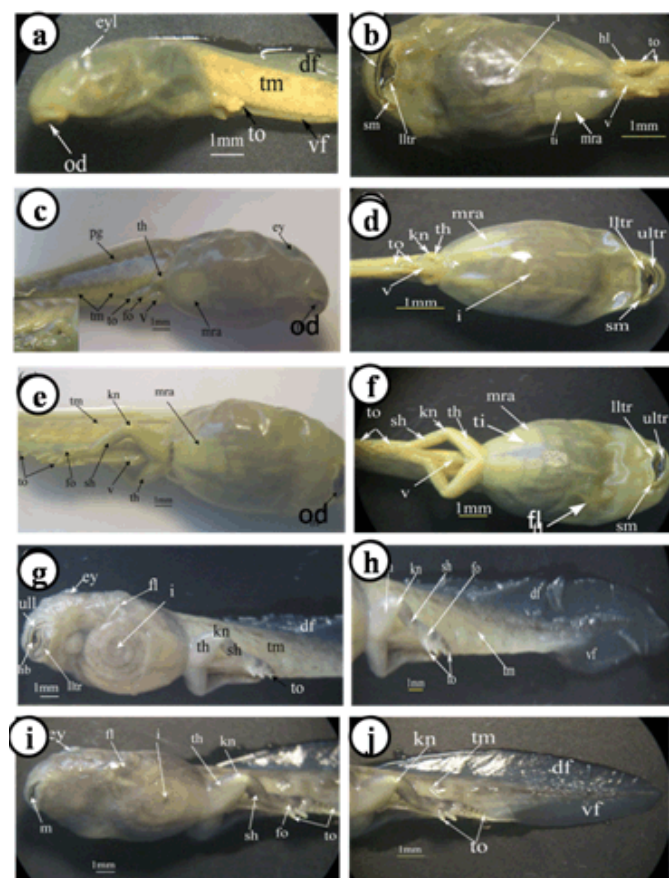


Figure 4: (a & b) Developmental embryonic stages of *Bufo regularis*, Lateral and ventral view of stage 55 showing, third indentation behind the third toe. (c & d) Lateral and ventral view of stage 56, in situ showing forth indentation and appearance of the fifth toe. (e & f) Lateral and ventral view of stage 57 showing well differentiated hind limb and elongation of the forth toe. And well differentiated *M. rectus abdominis* with its five compartments. (g & h) Lateral and ventral view of anterior and posterior portion of stage 58. (i & j) Lateral and ventral view of anterior and posterior portion of stage 58b showing, regression and darkening of the caudal fin, and regression of the oral disc, df = dorsal fin, ey = eye, eyl = eye lens, fl = forelimb, fo = foot, hb = hind limb, i = intestine, kn = knee, lltr = lower lip of teeth rows, m = mouth, mra = muscle rectus abdominis, od = oral disc, pg = pigments, sh = shank, sm = sub marginal papillae, th = thighbone, ti = tendinous intersections, tm = tail muscle, to = toes, ultr = upper lip of teeth rows, v = vent, vf = ventral fin and scale bar = 1 mm.

in the ventral side of the toadlet was clearly detected. *M. rectus* abdominis disappeared under the thick skin of the toadlet. Well-developed two pairs of legs; forelimb had four fingers and hind limb was longer than the forelimb and had five toes (Figure 5e&f and 6o). The development of forelimbs in different developmental stages of *Bufo regularis* was given in (Figure 7).

DISCUSSION

In the present study, developmental changes in external morphology characters and morphometric measurements were examined for the larval period of *Bufo regularis*. A developmental table consisting of 13 developmental stages from starting of feeding to complication of tail absorption every third day was recorded.

To be useful, a staging table must be based on character states whose onset of development can be detected with minimal ambiguity, whose presence among individuals is stable, and, preferably, whose characteristics are quantifiable [6].

Gosner [10] suggested a generalized table of 46 stages covering the period from fertilization of the egg to completion of metamorphosis and this table has been

used for the comparison of developmental stages of anurans world over.

Sedra and Michael [20] had established a set of closely spaced developmental stages for the Egyptian toad *Bufo regularis*. These include stages numbered 1–66 which begin with the zygote stage and end with the newly metamorphosed individuals. We had used Sedra and

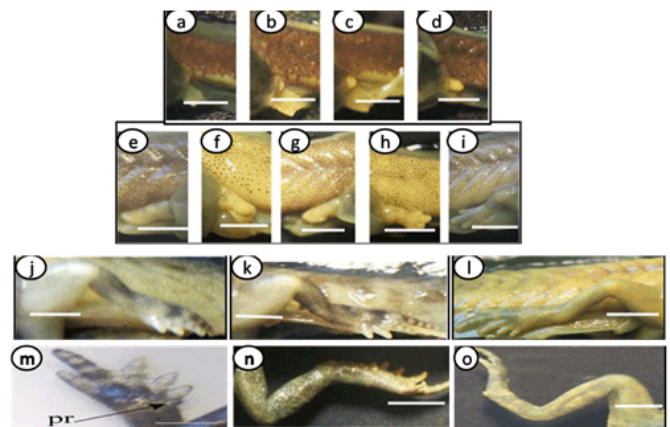


Figure 6: Development of hind limb in different developmental stage of *B. regularis* Showing, dorsal views of the developing hind limbs (a-i, k, l), ventral views (j, m, n), lateral view (o). (a & b) Stage 46, (c) Stage 48, (d) Stage 49, (e) Stage 52, (f) Stage 53, (g) Stage 54, (h) Stage 55, (i) Stage 56, (j) Stage 57, (k) Stage 58, (l) Stage 59, (m) Stage 60, (n) Stage 64, (o) Stage 66, Prehallux (pr) and scale bar = 1 mm.

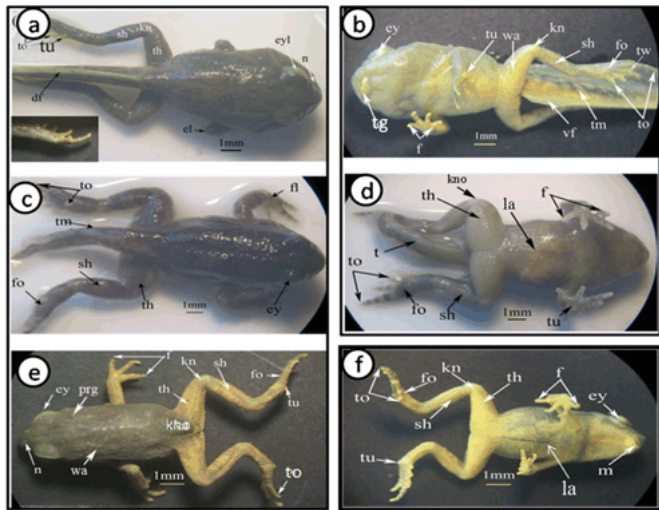


Figure 5: Developmental embryonic stages of *Bufo regularis* (a & b) Dorsal and ventral view of stage 59 showing emergence of the forelimbs appearance of warts on back skin, fallen of the oral disc. (c & d) Dorsal and ventral view of stage 64 showing regressing of the tail. (e & f) Dorsal and ventral view of stage 66, abv = abdominal vein, bc = blood capillaries, df = dorsal fin, ey = eye, eyl = eye lens, f = finger, fl = forelimb, fo = foot, hb = hind limb, i = intestine, kn = knee, kno = tail knob, la = linea alba, lltr = lower lip of teeth rows, m = mouth, mra = muscle rectus abdomens, n = nose, od = oral disc, op = operculum, pg = pigments, prg = parotoid gland, sh = shank, sm = sub marginal papillae, t = tail, th = thigh, tm = tail muscle, tongue (tg), to = toes, tu = tubercle, ultr = upper lip of teeth rows, v = vent, vf = ventral fin, wa = warts and scale bar = 1 mm.

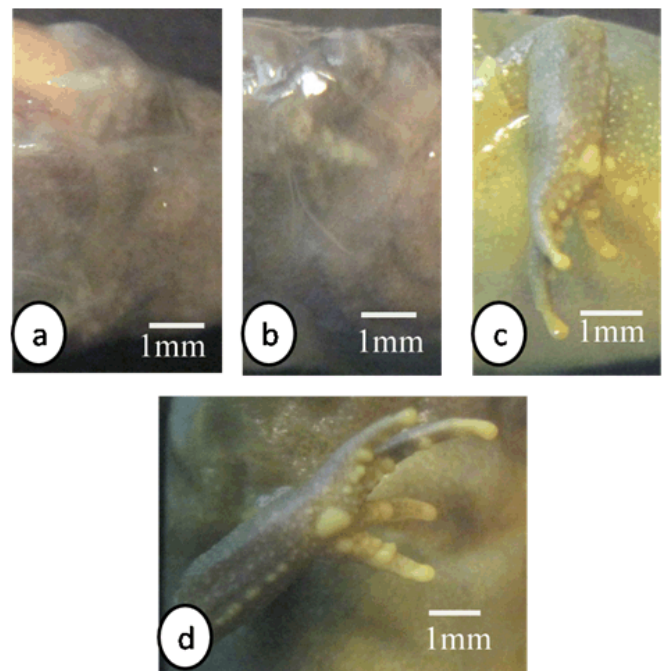


Figure 7: Development of forelimbs in different developmental stage of *B. regularis* showing, the emergence of the well developed forelimbs with well developed fingers and subarticular tubercles. (a) Stage 58a, (b) Stage 58b (c) Stage 59, (d) Stage 64 and scale bar = 1 mm.

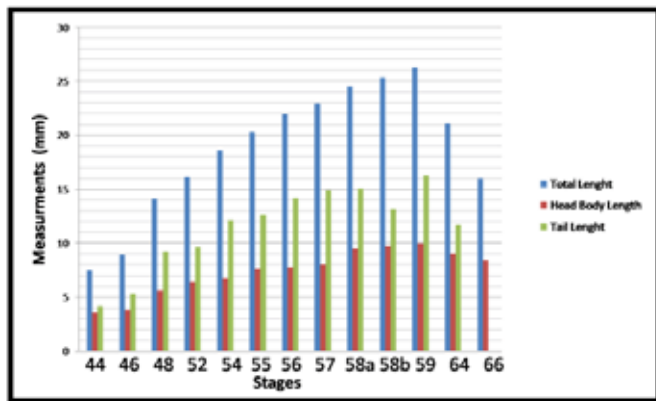


Figure 8: Relation between total length, head body length and tail length in developmental stages of Egyptian toad *Bufo regularis*.

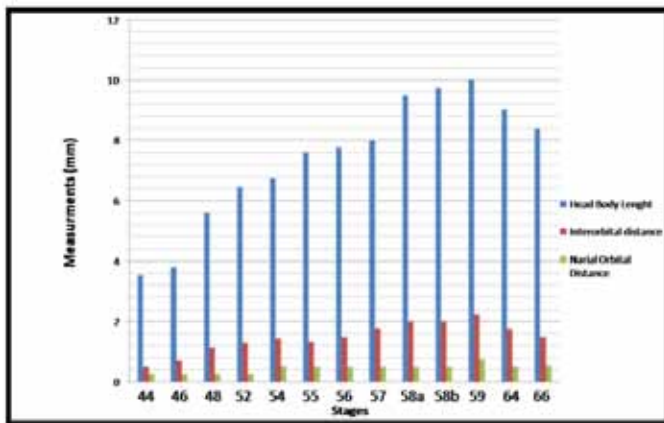


Figure 9: Relation between head body length, interorbital distance and distance between nasal and orbital in developmental stages of Egyptian toad *Bufo regularis*.

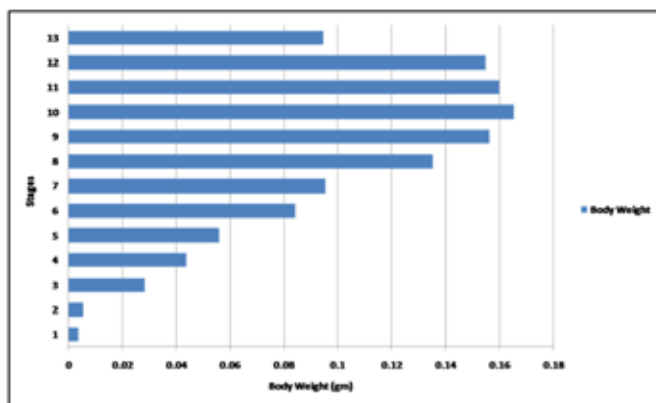


Figure 10: The variability of body weight in developmental stages of *Bufo regularis*. (1) Stage 44, (2) Stage 46, (3) Stage 48, (4) Stage 52, (5) Stage 54, (6) Stage 55, (7) Stage 56, (8) Stage 57, (9) Stage 58a, (10) Stage 58b, (11) Stage 59, (12) Stage 64, (13) Stage 66.

Michael [20] staging table for identifying the stages of development of the studied tadpoles for its simplicity and general applicability.

Morphology

In this section, we discuss the specific portions of the staging table in which we determined that a modification to [20] was necessary. Also, we include the timing of occurrence of certain developmental events and compare them with similar data from the other literatures.

In the present study, we recorded that feeding of *B. regularis* tadpoles, began at stage 44. While in previous studies by Sedra and Michael [20] on the same species noticed that at the beginning of this stage the tadpoles began route for start feeding. Gosner [10] considered the stage 25 start feeding in *Bufo valliceps*. In *Scaphiopus bombifrons* the feeding began at stage 24 [26] while in *Scaphiopus intermontanus* feeding began in either stage 24 or 25A [6], but in *Microhyla ornate* tadpole feeding began at stage 28 [27]. In *Rana leptoglossa* feeding began at stage 25 [4]. Hall and Larsen [6] declared that the larvae were able to begin feeding once their jaws had begun to keratinize.

Sedra and Michael [20] recorded that the staging criteria associated with the development of the hind limbs which first noticeable in *B. regularis* during stage 46. These results were similar to the development of the hind limb bud in the Egyptian toad (*B. regularis*) tadpole in the present work.

According to Gosner's table [10] in *Bufo valliceps* [10, 28] and in *Rana pipiens* [28] marked the initial presence of the hind limb bud as subsequent to the completion of opercular development in stage 26. In contrast, in *Rana sylvatica* [29], the bud is identifiable at the time when operculum begins development (stage 23), however, [6] noticed the first appearance of the hind limb bud in *S. intermontanus* during stage 24 (after opercular development had already progressed significantly but prior to completion). Similarly, Trowbridge [26] noted the first development of the hind limb bud during stage 23 in *Scaphiopus bombifrons* [6] described the first appearance of the hind limb bud in *Scaphiopus intermontanus* tadpole as an opaque swelling of tissue posterior to the abdomen and dorsal to the cloacal tailpiece. This description was in accordance with the present work.

In our study in *B. regularis* foot paddle formation began at stage 52, when hind limb length reached one and a half times its diameter [10] had defined the foot paddle from the point when limb bud length reaches two times its diameter but formation of the foot paddle had not yet begun, and he recorded that the limb bud show as essentially bullet shaped prior to foot paddle formation (probably based on the description of stage 30 for *Bufo valliceps* by Limbaugh and Volpe [28], for *Rana leptoglossa* by Saha and Gupta [4], and for *Microhyla*

ornata and *Uperodon globulosus* by Narzary and Bordoloi [18] confirmed the results of Gosner [10]. Also, they reported that stage 31 was the foot paddle shaped stage. [6] had been observed in *S. intermontanus* foot paddle formation began at about the point hind limb bud length was only slightly greater than its diameter. Our result was in agreement of [20] where they stated that, foot paddle formation in *B. regularis* occurs prior to the limb bud achieving a length of two times its diameter, but after it is one and a half times its diameter.

In the present investigation, toe development in the hind limb of *B. regularis* tadpoles takes place from stage 52–56, where these stages takes degrees of toe development. Gosner and Taylor stated that a combination of degree of toe separation was recorded from stage 36–40 [10, 30] but, in *Rana leptoglossa* it had been observed that the toe differentiation occur during stages 31 to 39 [4].

In the present work, we observed first indentation between fifth and fourth toes in late stage 52. Whereas, the first indentation was noticed by Sedra and Michael [20] in stage 53 in the same species. The 2nd indentation was seen behind the fourth indentation which marks the third toe in stage 54, third indentation marks the 2nd toe at stage 55. The fourth indentation marks the 1st toe in stage 56. All five toes were visible and separated from each other. These results were similar to those described by Sedra and Micheal [20] in *B. regularis*. The same events were recorded by several authors [4, 18, 27] in *Microhyla ornata*, *Rana leptoglossa* and *Uperodon globulosus* respectively, where they observed the margin of the foot paddle became slightly intended which marked the prominences of the fifth and fourth toes at stage 32, second indentation marks the 3rd toe in stage 33, third indentation marks the 2nd toe at stage 34 and the fourth indentation marks the 1st toe at stage 35.

Landmarks between thigh, shank and foot with separation of all toes began at stage 56 and well differentiated at stage 57. At this stage well developed prehallux appeared as a small protuberance close to the base of the first toe. These results were similar to that described by Sedra and Micheal [20] at stages 56 and 57 in *B. regularis*.

In our results in stage 57, we observed with the separation of all toes the fourth toe is the longest. Web clearly developed between 2–3 and 3–4 toes. Instep carrying little tubercles some of which are enlarged at bases of the second, third, and fourth toes. These results are in agreement with those of Sedra and Michael [20]. The same results were recorded in stage 39–40 by several authors [4, 18, 27] in their studies.

Our staging for *B. regularis* recorded that both forelimbs emergence and external nares are opened in late stage 59, and these results were similar to [18, 27] where they had been observed that the forelimbs emerged at (stage 42) in *Microhyla ornata*, also, these results were similar to Gosner and McDiarmid et al. [10, 31] where

they observed that external nares open very late, after forelimbs emerge at Gosner's stage 41, and nares opened forming a rim by the nasal opening in Gosner's stage 42 [32].

In our results in stage 64, we recorded that the tail was decreased in size and the dorsal and ventral lobes of caudal fin are highly reduced and represented by shallow crests and these results were similar to stage 64 [20], also, similar to stage 44 of *Microhyla ornata* [27]. In *Rana leptoglossa*, [4] observed the tail resorption started at stage 43 with shrinking in dorsal and ventral fins. Some authors had been used the degree of tail resorption and emergence of the forelimbs as an indexes of progress in metamorphosis and they had been recorded that the tail shortening begins at about the time of forelimb emergence [7, 28]. Complete of the tail regression signals the end of metamorphosis in all tadpoles [6].

We observed the angle of the mouth reaching the posterior margin of the eye in stage 64 in *B. regularis*; this result was in agreement with Sedra and Michael [20]. This observation was recorded in stage 45 in *Bufo valliceps* and *Uperodon globulosus* by Gosner [10, 18] but in *S. intermontanus* it had been observed only in sexually mature individuals [6].

Studies on the mechanism of apoptotic cell death in the tail suggested that activation of collagenase might play an important role in the disappearance of the tail [15]. However, degradation of extracellular matrix proteins (ECM) induces proliferation, migration and differentiation of constituent cells during embryonic development [33, 34]. It has been reported that catalase inhibited programmed cell death of blastocytes and H₂O₂ stimulated apoptosis [35, 36]. Thus, in addition to collagenase, reactive oxygen species might play critical roles in the apoptosis of cells in the tail.

Our results in late stage 66, we observed completion of tail resorption and these results were similar to stage 66 [20] where they had been observed that the tail had nearly disappeared and is only represented by a minute knob which is no longer visible in a ventral aspect, and these results were similar to (stage 46) of *Microhyla ornata* and *Uperodon globulosus* [18, 27], similarly, the tail resorption was complete in stage 45 but still remained a triangular stub in *Rana leptoglossa* [4].

The stages of morphogenesis of the muscle rectus abdominis of the tadpoles *Bufo regularis* had been described from the stage of its appearance (stage 48) to the stage of its hidden under the thick skin of the tadpole at (stage 59) by Sedra and Michael [20]. In the present investigation this muscle of rectus abdominis appeared at stage 46 and obscured under the thick and opaque skin also at stage 59 (Figures 2, 4, and 5). The full developed stages of this muscle in the present investigation had been recorded from stage 56–58 but in Sedra and Michael [20] from stage 51–58. In the present work these muscles and the linea alba are well distinguished through the thin, pale toadlet skin (Figure 5f).

Table 1: Morphometric measurements of developmental stages of Egyptian tadpoles *Bufo regularis*. The basic statistics expressed as mean $\pm$ SD (range), N= 6

Stages Measurements**	Stage 44	Stage 46	Stage 48	Stage 52	Stage 54	Stage 55	Stage 56	Stage 57	Stage 58a	Stage 58b	Stage 59	Stage 64	Stage 66
TL(mm)1	7.5 $\pm$ 0 (7.5-7.5)	8.75 $\pm$ 0.37 (8-10)	14.41 $\pm$ 0.68 (12.25-15.5)	16.14 $\pm$ 0.31 (15.3-17.4)	18.64 $\pm$ 0.22 (17.9-19.2)	20.26 $\pm$ 0.10 (20-20.5)	21.95 $\pm$ 0.02 (21.9-22)	22.95 $\pm$ 0.02 (22.9-23)	24.15 $\pm$ 0.07 (24.3-24.75)	25.37 $\pm$ 0.05 (25.25-25.5)	26.25 $\pm$ 0.11 (26-26.5)	21.8 $\pm$ 0.05 (21-21.25)	15.95 $\pm$ 0.03 (15.8-16)
H-BL(mm)1	3.54 $\pm$ 0.04 (3.5-3.75)	3.73 $\pm$ 0.24 (3-4.25)	5.61 $\pm$ 0.13 (5.2-6)	6.45 $\pm$ 0.11 (6.55-7)	6.76 $\pm$ 0.07 (6.5-7)	7.62 $\pm$ 0.05 (7.5-7.75)	7.77 $\pm$ 0.01 (7.75-7.8)	8 $\pm$ 0 (8-8)	9.50 $\pm$ 0 (9.5-9.5)	9.75 $\pm$ 0.11 (9.5-10)	10 $\pm$ 0 (10-10)	9.04 $\pm$ 0.04 (9-9.25)	8.41 $\pm$ 0.05 (8.25-8.5)
BW(mm)1	2.04 $\pm$ 0.04 (2-2.25)	2.55 $\pm$ 0.16 (2-3)	3.45 $\pm$ 0.25 (2.75-4)	4.04 $\pm$ 0.15 (3.75-4.75)	4.64 $\pm$ 0.10 (4.4-5)	4.8 $\pm$ 0.08 (4.6-5)	4.95 $\pm$ 0.04 (4.75-5)	6.12 $\pm$ 0.05 (6-6.25)	6.63 $\pm$ 0.06 (6.5-6.8)	6.64 $\pm$ 0.06 (6.5-6.8)	6.08 $\pm$ 0.28 (5.25-6.8)	6.28 $\pm$ 0.07 (6-6.5)	4.11 $\pm$ 0.04 (4-4.2)
BH(mm)1	1.76 $\pm$ 0.10 (1.5-2.1)	2.11 $\pm$ 0.09 (1.75-2.25)	3.29 $\pm$ 0.14 (3-3.75)	3.37 $\pm$ 0.16 (2.75-4)	3.57 $\pm$ 0.16 (3-4)	3.88 $\pm$ 0.14 (3.5-4.3)	4.20 $\pm$ 0.04 (4-4.25)	5.2 $\pm$ 0 (5.2-5.2)	4.62 $\pm$ 0.12 (4.5-5.25)	5.05 $\pm$ 0.18 (4.5-5.5)	4.77 $\pm$ 0.19 (4.25-5.5)	4.59 $\pm$ 0.13 (4-4.5)	4.11 $\pm$ 0.07 (4-4.4)
TAL(mm)1	4 $\pm$ 0.12 (3.5-4.25)	5.27 $\pm$ 0.26 (4.25-6)	8.3 $\pm$ 0.05 (8.25-8.5)	9.62 $\pm$ 0.28 (9-10.75)	12.15 $\pm$ 0.12 (11.75-12.5)	12.62 $\pm$ 0.05 (12.5-12.75)	14.2 $\pm$ 0.02 (14.15-14.25)	14.95 $\pm$ 0.02 (14.15-15)	15.05 $\pm$ 0.07 (14.8-15.25)	15.62 $\pm$ 0.05 (15.5-15.75)	16.25 $\pm$ 0.11 (16-16.5)	11.66 $\pm$ 0.21 (11-12)	0
TH(mm)1	0.74 $\pm$ 0.06 (0.5-1)	1.44 $\pm$ 0.08 (1.25-1.7)	2.88 $\pm$ 0.12 (2-2.75)	2.66 $\pm$ 0.12 (2.25-3)	2.72 $\pm$ 0.21 (2.25-3.75)	2.75 $\pm$ 0.26 (2.25-3.75)	2.85 $\pm$ 0.06 (2.6-3)	3.67 $\pm$ 0.04 (3.5-3.75)	4.41 $\pm$ 0.08 (4-4.6)	4.31 $\pm$ 0.19 (3.5-4.7)	4.22 $\pm$ 0.20 (3.25-4.6)	1.75 $\pm$ 0.11 (1.5-2)	0
TMH(mm)1	0.33 $\pm$ 0.05 (0.25-0.5)	0.48 $\pm$ 0.02 (0.4-0.5)	0.6 $\pm$ 0.1 (0.5-1)	1.15 $\pm$ 0.04 (1-1.25)	1.43 $\pm$ 0.03 (1.3-1.5)	1.53 $\pm$ 0.02 (1.5-1.6)	1.58 $\pm$ 0.05 (1.5-1.75)	2.64 $\pm$ 0.05 (2.5-2.78)	2.7 $\pm$ 0.04 (2.6-2.8)	2.43 $\pm$ 0.033 (2.3-2.5)	1.83 $\pm$ 0.05 (1.7-2)	1.16 $\pm$ 0.05 (1-1.25)	0
ED(mm)1	0.25 $\pm$ 0 (0.25-0.25)	0.37 $\pm$ 0.05 (0.25-0.5)	0.6 $\pm$ 0.04 (0.5-0.7)	0.76 $\pm$ 0.01 (0.75-0.8)	0.85 $\pm$ 0.02 (0.8-0.9)	0.9 $\pm$ 0.04 (0.8-1)	1 $\pm$ 0 (1-1)	1.145 $\pm$ 0.05 (1-1.25)	1.37 $\pm$ 0.05 (1.25-1.5)	1.5 $\pm$ 0 (1.5-1.5)	1.8 $\pm$ 0.06 (1.6-1.9)	2.25 $\pm$ 0.11 (2-2.25)	1.75 $\pm$ 0.11 (1.5-2)

Table 1: (Continued)

ND(mm)1	0.125±0.011 (0.1-0.15)	0.18±0.17 (0.5-0.8)	1±0 (1-1)	0.25±0 (0.25-0.25)	1.42±0.03 (1.3-1.5)	0.75±0 (0.75-0.75)	0.33±0.02 (0.3-0.4)	0.35±0.02 (0.3-0.4)	0.43±0.02 (0.4-0.5)	0.42±0.06 (0.3-0.6)	0.27±0.01 (0.25-0.34)	1.4±0.04 (1.3-1.5)	1.33±0.05 (1.25-1.5)
IOD(mm)1	0.5±0 (0.5-0.5)	0.72±0.04 (0.5-0.8)	1.12±0.05 (1-1.25)	1.29±0.15 (1-2)	1.43±0.03 (1.3-1.5)	1.33±0.05 (1.25-1.5)	1.5±0 (1.5-1.5)	1.75±0.00 (1.75-1.8)	2±0 (2-2)	2±0 (2-2)	2.25±0 (2.25-2.25)	1.75±0 (1.75-1.75)	2±0 (2-2)
IND(mm)1	0.25±0 (0.25-0.25)	0.33±0.03 (0.2-0.4)	0.65±0.03 (0.6-0.75)	0.74±0.08 (0.5-1.1)	0.93±0.04 (0.8-1.1)	0.79±0.04 (0.75-1)	0.9±0.04 (0.8-1)	1.1±0.16 (0.8-1.9)	1.01±0.01 (1-1.1)	1.05±0.03 (1-1.2)	0.80±0.09 (0.5-1)	1.13±0.17 (0.75-1.8)	0.53±0.02 (0.5-0.6)
NOD(mm)1	0.25±0 (0.25-0.25)	0.25±0 (0.25-0.25)	0.25±0 (0.25-0.25)	0.45±0.04 (0.25-0.5)	0.52±0.12 (0.25-0.8)	0.5±0 (0.5-0.5)	0.5±0 (0.5-0.5)	0.5±0 (0.5-0.5)	0.5±0 (0.5-0.5)	0.5±0 (0.5-0.5)	0.75±0 (0.75-0.75)	0.5±0 (0.5-0.5)	0.75±0 (0.75-0.75)
HLL(mm)1	0	0.12±0.002 (0.12-0.13)	0.53±0.02 (0.5-0.6)	0.56±0.02 (0.5-0.6)	0.72±0.03 (0.6-0.75)	1.16±0.10 (0.75-1.5)	1.7±0.03 (1.6-1.75)	1.9±0.05 (1.75-2)	5.75±0.11 (5.5-6)	7.291±0.02 (7.25-7.4)	9.08±0.5 (9.75-10)	11.94±0.04 (11.75-12)	7.91±0.26 (7.5-8.75)
FLL(mm)1	*	*	*	*	*	*	*	*	*	*	6.37±0.05 (6.25-6.5)	6.5±0.02 (6.5-6.6)	4.25±0.11 (4-4.5)
BWE (g)2	0.003±0.0003 (0.003-0.005)	0.005±0.0004 (0.0043-0.0068)	0.028±0.0017 (0.0249-0.0368)	0.043±0.0006 (0.041-0.046)	0.056±0.0003 (0.054-0.057)	0.084±0.001 (0.081-0.087)	0.095±0.005 (0.094-0.097)	0.135±0.006 (0.133-0.138)	0.165±0.008 (0.163-0.168)	0.156±0.005 (0.154-0.158)	0.160±0.002 (0.15-0.16)	0.155±0.0005 (0.153-0.156)	0.094±0.0004 (0.093-0.096)

* This structure will be disappeared

** Total Length (TL), Head- Body Length (H-BL), Body Wide, (BW), Body Height (BH), Tail Length (TAL), Tail Height (TH), Tail Muscle Height (TMH), Eye Diameter (ED), Nostriell Diameter (ND), Interorbital Distance (IOD), Internariel Distance (IND), Distance between Nariel and orbital (NOD), Hind Limb Length (HLL), Fore Limb Length (FLL).

o This structure not appeared in this stage

1 Isometric growth and 2 Allometric growth

Table 2: Personal correlation of morphometric measurements of *Bufo regularis* tadpoles.

	STAGES	TL	HBL	BW	BH	TAL	TH	MTH	ED	ND	IOD	IND	NED	HLL	FLL	BWE
STAGES	1															
TL	.721**	1														
HBL	.897**	.933**	1													
BW	.754**	.941**	.917**	1												
BH	.799**	.904**	.905**	.923**	1											
TAL	.227*	.775**	.560**	.702**	.582**	1										
TH	0.215	.775**	.575**	.730**	.628**	.842**	1									
MTH	.376**	.859**	.684**	.816**	.717**	.866**	.937**	1								
ED	.936**	.672**	.844**	.725**	.731**	.251*	0.185	.324**	1							
ND	.572**	.716**	.698**	.791**	.736**	.469**	.554**	.595**	.517**	1						
IOD	.893**	.856**	.941**	.843**	.874**	.433**	.526**	.620**	.802**	.622**	1					
IND	.494**	.700**	.641**	.776**	.710**	.582**	.581**	.614**	.504**	.598**	.586**	1				
NED	.741**	.601**	.703**	.541**	.622**	0.165	.229*	.328**	.666**	.333**	.751**	.311**	1			
HLL	.894**	.569**	.770**	.646**	.613**	0.168	0.143	.281*	.922**	.477**	.749**	.399**	.592**	1		
FLL	.698**	.277*	.484**	.315**	.332**	-0.016	-0.153	-0.011	.783**	0.116	.499**	0.156	.536**	.850**	1	
BWE	.870**	.918**	.953**	.947**	.905**	.597**	.615**	.752**	.826**	.735**	.893**	.668**	.613**	.790**	.474**	1

* Significant at $P < 0.05$

** Significant at $P < 0.01$

Morphometric measurements

In this section, we reported morphometric data on *B. regularis* tadpoles from beginning of feeding every three days to complication of metamorphosis (absorption of the tail), we measured the following variables: BH, BW, ED, FLL, H-BL, HLL, IND, IOD, ND, NOD, TAL, TH, TL, TMH, and BWE, as given in Tables [1], those measurements considered as biomarkers for morphology growth of the tadpole body.

It had been reported that the morphometric characters of tadpoles were widely used in interspecific comparison and in taxonomic studies to understand these characters and their morphology in the adult phase [37]. Also, the different morphometric variables show specific increasing or decreasing patterns in different stages [38].

Our results showed fluctuation in the TL, H-BL, BH, BW, TAL, TH and TMH, during stages 44–66, and this similar to Dey et al. and Grosjean et al. [38, 39].

Previous studies have demonstrated that in the presence of predators, tail heights of tadpole become taller and overall body length become shorter [19, 40, 41]. Also found that tadpoles with this increased tail height survived predation 25% more than those without increased tail heights. Similarly, Vences et al. [42] suggested that taller tail heights may be more beneficial in larger bodies of water where longer daily distances are moved than in smaller bodies of water. This is possibly due to thermoregulatory functions and possibly feeding

activity. Thus the greater tail heights of *Rana clamitans* may be a result of their ability to exploit more habitat types as opposed to *R. okaloosa* that only selects small bogs and seeps in Northwestern Florida [43–45].

Eye diameter and the inter-orbital distance was increased with age from (stage 44 to stage 64), then decreased in the late stage (stage 66), and this similar to *Rana alticola* [39–46] and these may be due to the change of the eye position from dorsal to dorsolateral position.

Kruger et al. [47] suggested that dorsal eyes may face dorsally (upward), dorsolaterally, or laterally, while lateral eyes always face laterally [48]. No part of the cornea of the eye is in the dorsal silhouette when the eyes are dorsally situated. Dorsal eyes occur in benthic tadpoles in both lentic and lotic systems, while lateral eyes are most common in lentic tadpoles that spend considerable time in the water column.

Also, we recorded fluctuation in the nostril diameter along the developmental stages, and this may be due to the development of respiratory system as was gills then become lungs.

The interorbital distance and internarial distance also showed fluctuation along different developmental stages and this was in accordance with results of [38].

Our results showed significantly increased in hind limb length with age from (stage 46–64), then decreased in the late stage (stage 66), also, forelimb length slightly increased in stage (59 and 64) then decreased in stage 66. These results were in contrast with [38], where they

recorded that the length of fore limbs continuously increased with age until tadpoles reached adult age in *Microhyla ornate*.

Table 1, and Figure 10 showed increased in the body weight from stages 44 to 58b, then decreased in stage 59, 64, and 66, and this was similar to [49]. In contrast, [2] observed fluctuation in the body mass through development of *Rhinella icterica*.

Decreasing in body weight was noticed from stage 59, 64 and 66 owing to the tadpoles stopped eating during the metamorphic process [20] and due to regression of the tail of the tadpoles *Bufo regularis* during this period and also the gradually fallen of the oral disc at this period.

CONCLUSION

Using morphology and morphometry indicated many differences between the different developmental stages and this study may be considered as morphological key for the different developmental stages of the Egyptian toad tadpoles (*Bufo regularis*).

Acknowledgements

This research funded by Assiut university and the Ministry Higher Education from Libyan Government.

Author Contributions

Alaa El-Din H. Sayed – Substantial contributions to conception and design, Acquisition of data, Analysis and interpretation of data, Drafting the article, Revising it critically for important intellectual content, Final approval of the version to be published

Ekbal T. Wassif – Analysis and interpretation of data, Revising it critically for important intellectual content, Final approval of the version to be published

Afaf I. Elballouz – Analysis and interpretation of data, Revising it critically for important intellectual content, Final approval of the version to be published

Guarantor

The corresponding author is the guarantor of submission.

Conflict of Interest

Authors declare no conflict of interest.

Copyright

© 2016 Alaa El-Din H. Sayed et al. This article is distributed under the terms of Creative Commons Attribution License which permits unrestricted use, distribution and reproduction in any medium provided the original author(s) and original publisher are properly credited. Please see the copyright policy on the journal website for more information.

REFERENCE

1. Saad AH, Aziz AA, Yahie A, El-Ghareeb A. Programmed cell death in the liver of different species of anuran amphibians during metamorphosis. Australian Journal of Basic and Applied Sciences 2009;3(4):4644–55.
2. Lima MS, Pederassi J. Morphometrics and ratio of body proportionality of tadpoles of *Rhinella icterica* (Anura, Bufonidae) at different developmental stages. Braz J Biol 2012 Aug;72(3):623–9.
3. Wright ML, Richardson SE, Bigos JM. The fat body of bullfrog (*Lithobates catesbeianus*) tadpoles during metamorphosis: changes in mass, histology, and melatonin content and effect of food deprivation. Comp Biochem Physiol A Mol Integr Physiol 2011 Dec;160(4):498–503.
4. Saha BK, Gupta BBP. The development and Metamorphosis of an endangered frog, *Rana leptoglossa* (Cope, 1868). International Journal of Advanced Biological Research 2011;1(1):67–76.
5. Fox H. Amphibia Morphogenesis. Clifton, New Jersey: Humana press; 1984.
6. Hall JA, Larsen JH Jr. Postembryonic ontogeny of the spadefoot toad, *Scaphiopus intermontanus* (Anura: Pelobatidae): skeletal morphology. J Morphol 1998 Nov;238(2):179–244.
7. Etkin W. Growth and resorption phenomena in anuran metamorphosis I. Physiol Zool 1932;5:275–300.
8. Etkin W. Metamorphosis. In: Moore JA ed. Physiology of the Amphibia. New York: Academic Press; 1964.
9. Etkin W. Hormonal control of amphibian metamorphosis. In: Etkin W, Gilbert LI eds. Metamorphosis: a Problem in Developmental Biology. New York: Appleton-Century-Crofts; 1968. p. 313–48.
10. Gosner F. A simplified table for staging anuran embryos and larvae with notes on identification. Herpetological Conservation and Biology 1960;116:183–90.
11. Nodzenski E, Inger RF. Uncoupling of relative structure changes in metamorphosing torrent dwelling tadpoles. Copeia 1990;4:1047–54.
12. Huang TB, Brown DB. Prolactin is not a juvenile hormone in *Xenopus laevis* metamorphosis. PNAS 2000;97:195–9.
13. Furrow JD, Neff ES. A developmental switch induced by thyroid hormone: *Xenopus laevis* metamorphosis. Trends Endocrinol Metab 2006 Mar;17(2):40–7.
14. Page RB, Voss SR, Samuels AK, Smith JJ, Putta S, Beachy CK. Effect of thyroid hormone concentration on the transcriptional response underlying induced metamorphosis in the Mexican axolotl (*Ambystoma*). BMC Genomics 2008 Feb 11;9:78.
15. Dodd MHI, Dodd JM. Physiology of the Amphibia. In: Lofts B ed. The biology of metamorphosis. New York: Academic Press; 1976. P. 467–599.
16. Takada M, Kasai M. Prolactin increases open-channel density of epithelial Na⁺ channel in adult frog skin. J Exp Biol 2003 Apr;206(Pt 8):1319–23.
17. Wilbur HM, Collins JP. Ecological Aspects of Amphibian Metamorphosis: Nonnormal distributions of competitive ability reflect selection

- for facultative metamorphosis. *Science* 1973 Dec 28;182(4119):1305–14.
18. Narzary J, Bordoloi S. Study of normal development and external morphology of tadpoles of *Microhyla ornata* and *Uperodon globulosus* of the family microhylidae (Amphibia: Anura) from North East India. *International Journal of Advance Biological Research* 2013;3(1):61–73.
19. Periestley AS, Gorman TA, Haas CA. Comparative Morphology and Identification of Florida Bog Frog and Bronze Frog Tadpoles. *Biological Science* 2010;73(1):20–6.
20. Sedra SN, Micheal MI. Normal table of the Egyptian toad, *Bufo regularis* Reuss, with an addendum on the standardization of the stages considered in previous publications. *Cesk Morfol* 1961;9:333–51.
21. Lamotte M, Zuber-vogeli M. Contribution a l'étude des batraciens de TOuest African II. Le development larvaire de *Bufo regularis* Reuss, de *Rana occidentalis* Günther et de *Rana crassipes* (Buch. et peters.). *Bull Inst franc Afr noire* 1954;16(3):940.
22. Sayed AH, Elballouz AI, Wassif ET. Histological and Histochemical Studies on the Early Developmental Stages of the Egyptian Toad *Bufo regularis* Reuss. *Open Journal of Animal Sciences* 2015;5:142–56.
23. Sayed AH, Elballouz AI, Wassif ET. Molecular Characterization on the Early Developmental Stages of the Egyptian Toad *Bufo regularis* Reuss. *Open Journal of Genetics* 2014;4:343–54.
24. Altig R. A primer for the morphology of anuran tadpoles. *Herpetological Conservation and Biology* 2007;2(1):71–4.
25. SPSS. SPSS for Windows. Headquarters, Chicago: SPSS Inc; 1998.
26. Trowbridge MS. Studies on the normal development of *Scaphiopus bombifrons* Cope II. The latter embryonic and larval periods. *Trans Am Microsc Soc* 1942;61:66–83.
27. Shimizu S, Ota H. Normal development of *Microhyla ornata*: The First Description of the Complete Embryonic and Larval Stages for the Microhylid Frog (Amphibia: Anura). *Current Herpetology* 2003;22:73–90.
28. Limbaugh BA, Volpe EP. Early development of the Gulf Coast toad, *Bufo valliceps* Wiegmann. *Am Mus Novit* 1957;1842:1–32.
29. Pollister AW, Moore JA. Tables for the normal development of *Rana sylvatica*. *Anat Rec* 1937;68:489–96.
30. Taylor AC, Kollros JJ. Stages in the normal development of *Rana pipiens* larvae. *Anat Rec* 1946 Jan;94:7–13.
31. McDiarmid RW, Altig R. Research: materials and techniques. In: McDiarmid RW, Altig R eds. *Tadpoles: The Biology of Anuran Larvae*. Chicago: The University of Chicago press; 1999.
32. Bowatte G, Meegaskumbra M. Morphology and ecology of *Microhyla rubra* (Anura: Microhylidae) tadpoles from Sri Lanka. *Amphibia and Reptile Conservation* 2011;5(2):22–32.
33. Gross J. How tadpoles lose their tails. *J Invest Dermatol* 1966 Oct;47(4):274–7.
34. Alexander CM, Werb Z. Extracellular matrix degradation. In: Hay ED ed. *Cell Biology of Extracellular Matrix*. 2ed. New York: Plenum Press; 1991. P. 255–302.
35. Little GH, Flores A. Inhibition of programmed cell death by catalase and phenylalanine methyl ester. *Comp Biochem Physiol Comp Physiol* 1993 May;105(1):79–83.
36. Pierce GB, Parchment RE, Lewellyn AL. Hydrogen peroxide as a mediator of programmed cell death in the blastocyst. *Differentiation* 1991 Apr;46(3):181–6.
37. Grosjean S. The choice of external morphological characters and developmental stages for tadpole-based anuran taxonomy: a case study in *Rana* (*Sylvirana*) *nigrovittata* (Blyth, 1855) (Amphibia, Anura, Ranidae). *Contributions to zoology* 2005;74(1/2):61–76.
38. Dey M, Gupta A. A note on morphometry of tadpole and adult of *Microhyla ornata* (Dumeril & Bibron) (Anura: Microhylidae). *Zoos' Print Journal* 2002;17(12):951–3.
39. Grosjean S, Pérez M, Ohler A. Morphology and buccopharyngeal anatomy of the tadpole of *Rana* (*Nasirava*) *alticola* (Anura: Ranidae). *The Raffles Bulletin Of Zoology* 2003;51(1):101–7.
40. Kraft R, Escobar MM, Narro ML, et al. Phenotypes of *Drosophila* brain neurons in primary culture reveal a role for fascin in neurite shape and trajectory. *J Neurosci* 2006 Aug 23;26(34):8734–47.
41. Kraft PG, Wilson RS, Franklin CE. Predator-mediated phenotypic plasticity in tadpoles of the striped marsh frog, *Limnodynastes peronii*. *Austral Ecol* 2005;30:558–63.
42. Vences M, Puente M, Nieto S, Vieites DR. Phenotypic plasticity of anuran larvae: environmental variables influence body shape and oral morphology of *Rana temporaria* tadpoles. *J Zool London* 2002;257:155–62.
43. Moler PE. A new species of frog (Ranidae: *Rana*) from northwestern Florida. *Copei* 1985:379–83.
44. Moler PE. *Rana okaloosae* Moler Florida bog frog. *Catalogue of American Amphibians and Reptiles* 1993;561:1–3.
45. Bishop DC. Ecology and distribution of the Florida Bog Frog and Flatwoods Salamander on Eglin Air Force Base. Ph.D. Dissertation. Virginia Polytechnic Institute and State University, Blacksburg, VA. 2005.
46. Hoffmann H. Description of the previously unknown tadpole of *Hyalinobatrachium pulveratum* (Anura: Centrolenidae). *Rev Biol Trop* 2004 Mar;52(1):219–28.
47. Kruger DJD, Du Preez LH, Weldon C. Tadpole morphology of high altitude frogs from the Drakensberg mountains: North West University Yunibesiti Ya Bokone-Bophirima Noordwes-Universiteit; 2010.
48. McDiarmid RW, Altig R, eds. *Tadpoles: The Biology of Anuran Larvae*. Chicago: The University of Chicago Press; 1999.
49. Di Cerbo AR, Biancardi CM. Morphometric study on tadpoles of *Bombina variegata* (Linnaeus, 1758) (Anura; Bombinatoridae). *Acta Herpetologica* 2010;5(2):223–31.

Access full text article on
other devices



Access PDF of article on
other devices

